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(54) Title: ANTISENSE OLIGOS WHICH INTERFERE WITH mRNA CAP ACTIVITY AND INHIBIT TRANSLATION			
(57) Abstract			
<p>Compositions and methods for achieving therapeutic treatment of diseases and for regulating gene expression in biological experimental systems via interference with 5' capping of targeted nascent mRNAs are disclosed. Interference with capping is achieved in accordance with preferred embodiments utilizing antisense oligonucleotide compounds which are complementary to the targeted nascent RNA and prevent binding of capping enzyme either via steric interference or by chemical modification of the nascent mRNA.</p>			

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## ANTISENSE OLIGOS WHICH INTERFERE WITH mRNA CAP ACTIVITY AND INHIBIT TRANSLATION

The present application is a continuation-in-part application of United States Patent application 847,054, filed  
5 on April 3, 1992, which is a United States national phase application of PCT application US91/03606, filed May 22, 1991, which is a continuation-in-part of United States Patent application 527,559, filed May 23, 1990, now abandoned, all of which are incorporated herein by reference.

### 10 FIELD OF THE INVENTION

This invention is directed to methods for achieving therapeutic treatment of diseases and for regulating gene expression in biological experimental systems. The invention generally relates to the field of antisense oligonucleotide  
15 compounds in conjunction with the field of gene expression, especially protein expression, which is dependent upon the 5' cap of eucaryotic and viral RNAs.

### BACKGROUND OF THE INVENTION

Classical therapeutics has generally focused upon  
20 interactions with proteins in efforts to moderate their disease-causing or disease-potentiating functions. Such therapeutic approaches have many limitations. Recently, antisense oligonucleotide therapeutic and diagnostic approaches have been proposed.

25 Antisense methodology is the complementary hybridization of relatively short oligonucleotides to single-stranded mRNA or single-stranded DNA, or even double-

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stranded DNA, such that the normal, essential functions of these intracellular nucleic acids are disrupted. Hybridization is the sequence specific hydrogen bonding of oligonucleotides to RNA or single stranded DNA according to the rules of Watson-Crick base pairing. Such base pairs are said to be complementary to one another.

The events which disrupt nucleic acid function are discussed by Cohen in *Oligonucleotides: Antisense Inhibitors of Gene Expression*, CRC Press, Boca Raton FL, (1989) who proposes two possible types of terminating events. The first, hybridization arrest, denotes a terminating event in which the oligonucleotide inhibitor binds to the target nucleic acid and thus prevents, by simple steric hindrance, the binding of essential proteins, such as ribosomes or capping enzymes, to the nucleic acid. Methyl phosphonate oligonucleotides; P. S. Miller & P.O.P. Ts'O, *Anti-Cancer Drug Design*, Vol. 2, pp. 117-128 (1987); and  $\alpha$ -anomer oligonucleotides, Cohen J.S. ed., *Oligonucleotides: Antisense Inhibitors of Gene Expression*, CRC Press, Boca Raton FL (1989) are two of the most extensively studied antisense agents which are thought to disrupt nucleic acid function by hybridization arrest.

A second type of terminating event for antisense oligonucleotides involves enzymatic cleavage of the targeted RNA by intracellular RNase H. The oligonucleotide, which must be of the deoxyribo type, hybridizes with the targeted RNA and this duplex activates the RNase H enzyme to cleave the RNA strand, thus destroying the normal function of the RNA. Phosphorothioate oligonucleotides are a prominent example of an antisense agent which operates by this type of terminating event.

Considerable research is being directed to the application of oligonucleotides as antisense agents for therapeutic purposes. Applications of oligonucleotides as diagnostics, research reagents, and potential therapeutic agents require that the oligonucleotides be synthesized in large quantities, be transported across cell membranes or taken up by cells, appropriately hybridize to targeted RNA or DNA,

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and subsequently terminate or disrupt nucleic acid function. These critical functions depend on the initial stability of oligonucleotides towards nuclease degradation. O t h e r methodologies of action for oligonucleotide therapeutics and  
5 diagnostics have been sought in recent years and therapeutic and diagnostic compounds and compositions to exploit such methodologies are greatly desired. The present invention provides such new compounds and compositions together with methodologies for the use of antisense oligonucleotides in the  
10 diagnosis and treatment of disease in animals.

Most antisense approaches utilize antisense oligonucleotides targeted to the AUG, or, less often, the 5'-untranslated region, 3'-untranslated region, coding or other regions of mRNA. It is also known to use antisense  
15 oligonucleotides which target the 5' cap of mRNA to interfere with mRNA function, as disclosed in PCT publication WO 91/17755 (Baker), which is assigned to the same assignee as the present invention and incorporated herein by reference in its entirety. In the present invention, antisense oligonucleotides are used  
20 to prevent capping of the 5' end of mRNA in order to achieve the desired effect of interfering with mRNA function. Interference with mRNA function by preventing capping has been heretofore unknown.

Thus, there has been a long-felt need for  
25 oligonucleotides which interfere with mRNA function, are effective for therapeutic use and are possessed of no or few side effects. The present invention addresses this need.

#### OBJECTS OF THE INVENTION

It is a principal object of this invention to provide  
30 therapeutic methods and materials for the treatment of diseases through modulation of the activity of mRNA.

It is a further object of this invention to provide oligonucleotides for use in therapeutics.

A further object of this invention is to provide such  
35 oligonucleotides which are less likely to cause undesired or toxic side reactions or effects.

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Another object of this invention is to provide oligonucleotides that inhibit the maturation, stabilization, and/or translation of a selected mRNA.

Yet another object of this invention is to provide  
5 oligonucleotides that interfere with the capping of nascent mRNA.

A further object of this invention is to provide oligonucleotides that sterically or chemically interfere with the capping of nascent mRNA.

10 These and other objects will become apparent to persons of ordinary skill in the art from a review of the present specification and attendant claims.

#### SUMMARY OF THE INVENTION

This invention provides oligonucleotide compounds for  
15 inhibiting the capping of a nascent mRNA. The term "nascent mRNA" is commonly used to refer to mRNA molecules which are in the process of being transcribed (synthesized). The oligonucleotides are designed to be specifically hybridizable with the nascent mRNA to be inhibited and to interfere with the  
20 action or effect of capping enzyme upon the nascent mRNA. In preferred embodiments, the oligonucleotide compounds comprise modifications or functionalities which interfere with the capping reaction. The effect of the contacting of compounds of the invention with the targeted nascent mRNA is to impede or  
25 prevent the 5' capping of the nascent mRNA by enzymes such as guanylyl transferase such that it does not fully mature into mRNA. The incompletely matured mRNA either fails to be translated into protein altogether, or is translated to a diminished or impaired degree.

30 Since capping is generally a function of the 5' end of nascent mRNA, the specific hybridization desired is generally to a 5' portion of the nascent mRNA. The nascent mRNAs to be inhibited are preferably from about 20 to about 80 nucleotides in length. The compounds of the invention may  
35 operate by several mechanisms including steric interference with the 5' end of the nascent mRNA such that capping enzymes

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are prevented from performing their function. In a preferred embodiment, one or more nucleotides of the oligonucleotide of the invention "hang over" or "extend beyond" the 5' terminus of the nascent mRNA. By this is meant that the 3' portion of the  
5 oligonucleotide compound has nucleotide units in excess of those needed to hybridize with the 5' portion of the nascent mRNA.

Alternatively, the compounds, generally acting through their modifications or functionalities, may be such as to  
10 chemically modify the nascent mRNA in a manner inconsistent with its maturation. Such modification is preferably through the dephosphorylation of the 5' triphosphate or diphosphate of the nascent mRNA. The result of such a modification is generally a 5' monophosphate or hydroxyl group.

15 The methods of the invention inhibit capping of a nascent mRNA by interfering with the capping of a nascent mRNA, generally selected as being one which leads to a mature mRNA which is essential to the development or maintenance of a disease state. Generally the mRNA (and hence the nascent mRNA)  
20 selected is cell-, tissue- or disease-specific. The nascent mRNA is contacted with an oligonucleotide compound of the invention either directly, such as would be the case in certain diagnostic applications, or in cells, tissues or bodily fluids containing the nascent mRNA. The compound is present in an  
25 amount effective to diminish the mRNA activity or function. A preferred function to be interfered with is export of mRNA from cell nuclei. Another is splicing of the mRNA, and yet another is translation of the mRNA into protein.

#### DETAILED DESCRIPTION

30 It has been recognized that the majority of eucaryotic and viral small nuclear RNAs and messenger RNAs have a unique chemical structure at their 5' terminus which is required in varying degrees for their maturation, stability, and efficiency of translation. The general structural features are given in  
35 *J. Mol. Biol.* 99 519-547 (1975). The cap is a guanosine residue which is methylated at the nitrogen 7 position. It is

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joined to the penultimate 5' base of the RNA via a triphosphate linkage between the 5' hydroxyl groups of each residue. The 2' hydroxyls of the 5' terminal base(s) are methylated.

The cap structure is added to the 5' end of nascent mRNA transcripts during transcription in the nucleus. The term "nascent mRNA" is commonly used to refer to mRNA molecules which are in the process of being transcribed (synthesized). Capping occurs when the nascent mRNA, which emerges from the transcription machinery with the 5' end first, is from about 20 to 80 nucleotides in length. Coppola, J.A., Field, A.S., and Luse, D.S. (1983) *Proc. Natl. Acad. Sci.* 80:1251-1255. The cap site corresponds to the transcription initiation site of the mRNA. Capping is believed to occur in four steps: (1) hydrolysis of the 5'-terminal triphosphate of the nascent mRNA transcript to a diphosphate; (2) covalent attachment of a guanylate moiety to the 5' terminus via a 5'-to-5' triphosphate linkage; (3) methylation of the new guanine cap at the N7 position; and (4) methylation of the 2'-hydroxyl group of the first encoded nucleotide. These reactions are believed to be carried out by multifunctional protein complexes. At least three enzymatic activities are involved in the capping process: (1) an RNA triphosphate activity removes the 5'-terminal phosphate from the 5'-triphosphate of the nascent mRNA, (2) a guanylyltransferase activity transfers the guanine residue to the 5'-diphosphate of the nascent mRNA and, (3) a methyltransferase activity which methylates the guanine residue.

Once transcribed, the primary transcript from eucaryotic and certain viral genes must be processed due to the presence of intervening sequences within the coded regions of the transcript. The 5' cap has been shown to be necessary for processing of the primary transcripts to mature RNA molecules, specifically in the splicing reactions during which removal of the first intron occurs. See *Cell* 38 731-736 (1984). Absence of a 5' cap results in rapid degradation of the RNA in the nucleus and the cytoplasm as described in *Mol. Biol. Med.* 5 1-14 (1988) and *Cell* 32 681-694 (1983). Without the 5' cap the RNA is presumably accessible to 5' exonucleases. The 5' cap

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also plays an important role in transport of mRNA out of the nucleus. Hamm, J. and Mattaj, I.W. (1990) *Cell* 63:109. Finally, the majority of eucaryotic and many viral mRNAs studied to date require the 5' cap for the initiation of protein translation; See *Cell* 9 645-653 (1976); *Federation of Experimental Biologists Society Letter* 96 1-11 (1978); and *Prog. Nuc. Acid Res.* 35 173-207 (1988). There are also specific cap binding proteins which are components of the machinery required for the initiation of translation of a protein. See *Cell* 40 223-24 (1985); and *Prog. Nuc. Acid Res.* 35 173-207 (1988).

Based on the current understanding of the properties and function of the 5' cap structure of messenger RNA it is now believed that chemical or steric interference which prevents capping of nascent mRNA transcripts will preclude the transcripts from any pertinent processes, such as the production of an undesired protein. In general, the compositions necessary for modulating the activity of an RNA transcript in accordance with this invention may be regarded as oligonucleotides which sterically block or chemically modify the targeted nascent mRNA transcript to make the mRNA inaccessible to, or unrecognizable by, a capping enzyme, preferably an RNA triphosphatase or guanylyltransferase. The resulting uncapped transcript is unable to operate in one or more of its normal processes.

The oligonucleotides in accordance with this invention specifically hybridize with the selected nascent transcript in sufficient proximity to the 5' end to block or modify the site for 5' capping. The oligonucleotides of the present invention preferably have about 5 to about 50 nucleotide units. It is more preferred that such functionalities have from about 10 to 25 nucleotide units. As will be appreciated, a nucleotide unit is a base-sugar combination suitably bound to an adjacent nucleotide unit through phosphodiester or other bonds.

In the context of this invention, the term "oligonucleotide" refers to an oligomer or polymer of ribonucleic acid or deoxyribonucleic acid. This term includes oligomers consisting of naturally occurring bases, sugars and

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intersugar (backbone) linkages as well as oligomers having non-naturally occurring portions which function similarly. Such modified or substituted oligonucleotides are often preferred over native forms because of properties such as, for example, enhanced cellular uptake and increased stability in the presence of nucleases.

Specific examples of some preferred oligonucleotides envisioned for this invention may contain phosphorothioates, phosphotriesters, methyl phosphonates, short chain alkyl or cycloalkyl intersugar linkages or short chain heteroatomic or heterocyclic intersugar linkages. Most preferred are those with  $\text{CH}_2\text{-NH-O-CH}_2$ ,  $\text{CH}_2\text{-N(CH}_3\text{)-O-CH}_2$ ,  $\text{CH}_2\text{-O-N(CH}_3\text{)-CH}_2$ ,  $\text{CH}_2\text{-N(CH}_3\text{)-N(CH}_3\text{)-CH}_2$  and  $\text{O-N(CH}_3\text{)-CH}_2\text{-CH}_2$  backbones (where phosphodiester is  $\text{O-P-O-CH}_2$ ). Also preferred are oligonucleotides having morpholino backbone structures. Summerton, J.E. and Weller, D.D., U.S. Patent 5,034,506. In other preferred embodiments, such as the protein-nucleic acid (PNA) backbone, the phosphodiester backbone of the oligonucleotide may be replaced with a polyamide backbone, the bases being bound directly or indirectly to the aza nitrogen atoms of the polyamide backbone. P.E. Nielsen, M. Egholm, R.H. Berg, O. Buchardt, *Science* 1991, 254, 1497. Other preferred oligonucleotides may contain alkyl and halogen-substituted sugar moieties comprising one of the following at the 2' position: OH, SH,  $\text{SCH}_3$ , F, OCN,  $\text{O(CH}_2\text{)}_n\text{NH}_2$  or  $\text{O(CH}_2\text{)}_n\text{CH}_3$  where n is from 1 to about 10;  $\text{C}_1$  to  $\text{C}_{10}$  lower alkyl, substituted lower alkyl, alkaryl or aralkyl; Cl; Br; CN;  $\text{CF}_3$ ;  $\text{OCF}_3$ ; O-, S-, or N-alkyl; O-, S-, or N-alkenyl;  $\text{SOCH}_3$ ;  $\text{SO}_2\text{CH}_3$ ;  $\text{ONO}_2$ ;  $\text{NO}_2$ ;  $\text{N}_3$ ;  $\text{NH}_2$ ; heterocycloalkyl; heterocycloalkaryl; aminoalkylamino; polyalkylamino; substituted silyl; a fluorescein moiety, an RNA cleaving group; a conjugate; a reporter group; an intercalator; a group for improving the pharmacokinetic properties of an oligonucleotide; or a group for improving the pharmacodynamic properties of an oligonucleotide and other substituents having similar properties. In a preferred embodiment, oligonucleotides may contain one or more moieties capable of interfering with capping of the 5' end of a nascent mRNA target, either through

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steric interference or chemical modification of the nascent mRNA. In one preferred embodiment, the chemical modification is dephosphorylation of the 5'-triphosphate or 5'-diphosphate of the nascent mRNA, yielding a 5'-monophosphate or a hydroxyl group. The interfering moiety may preferably comprise an amino acid, peptide, nucleotide, polyamine, coordination complex of a metal, metal ion, or a macrocycle. In one preferred embodiment, the interfering portion comprises at least one nucleotide. The nucleotide or nucleotides of the interfering portion may be complementary to the nucleotides of the target mRNA according to the rules of Watson-Crick base pairing or may extend or overlap beyond the 5' end of the nascent mRNA.

Preferred examples of polyamines are triethylamine tetramine, diethylene triamine, and pentethyamine hexamine. Preferred examples of metal ions are transition metals such as cobalt, zinc, and copper; lanthanides such as europium and gadolinium, and divalent cations such as calcium and magnesium. Preferred examples of coordination complexes of metals are copper, zinc, cobalt or lanthanide complexes of bipyridine, orthophenanthroline, 12aneN3, diethylenetriamine pentaacetic acid, or N-[2-mercaptopropionyl]glycine. A preferred example of a heterocycle is the Lehn macrocycle, [24]ane N<sub>6</sub>O<sub>2</sub>. In a preferred embodiment of the invention, the interfering moiety is attached to the oligonucleotide at the 3' position. Oligonucleotides may also have sugar mimetics such as cyclobutyls or other carbocyclics in place of the pentofuranosyl group. Nucleotide units having nucleosides other than adenosine, cytidine, guanosine, thymidine and uridine may be used, for example inosine.

Chimeric oligonucleotides can also be employed; these molecules contain two or more chemically distinct regions, each comprising at least one nucleotide. These oligonucleotides typically contain a region of modified nucleotides that confer one or more beneficial properties (such as, for example, increased nuclease resistance, increased uptake into cells, increased binding affinity for the RNA target) and an

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unmodified region that retains the ability to direct RNase H cleavage.

The oligonucleotides used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis. Equipment for such synthesis is sold by several vendors including Applied Biosystems. Any other means for such synthesis may also be employed, however the actual synthesis of the oligonucleotides are well within the talents of the routineer. It is also well known to use similar techniques to prepare other oligonucleotides such as the phosphorothioates and alkylated derivatives.

While the present invention has been described with specificity in accordance with certain of its preferred embodiments, the following examples serve only to illustrate the invention and are not intended to limit the same.

#### EXAMPLES

##### Example 1

##### Oligonucleotide synthesis

Unmodified DNA oligonucleotides are synthesized on an automated DNA synthesizer (Applied Biosystems model 380B) using standard phosphoramidite chemistry with oxidation by iodine.  $\beta$ -cyanoethyldiisopropyl-phosphoramidites are purchased from Applied Biosystems (Foster City, CA). For phosphorothioate oligonucleotides, the standard oxidation bottle is replaced by a 0.2 M solution of 3H-1,2-benzodithiole-3-one 1,1-dioxide in acetonitrile for the stepwise thiation of the phosphite linkages. The thiation cycle wait step is increased to 68 seconds and is followed by the capping step.

2'-O-methyl phosphorothioate oligonucleotides are synthesized using 2'-O-methyl  $\beta$ -cyanoethyldiisopropyl-phosphoramidites (Chemgenes, Needham MA) and the standard cycle for unmodified oligonucleotides, except the wait step after pulse delivery of tetrazole and base is increased to 360 seconds.

After cleavage from the controlled pore glass column

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(Applied Biosystems) and deblocking in concentrated ammonium hydroxide at 55°C for 18 hours, the oligonucleotides are purified by precipitation twice out of 0.5 M NaCl with 2.5 volumes ethanol. Analytical gel electrophoresis is accomplished in 20% acrylamide, 8 M urea, 45 mM Tris-borate buffer, pH 7.0.

**Screen for evaluating oligonucleotide functional groups for ability to inhibit capping of nascent mRNAs**

The relative efficacy of various moieties and functional groups conjugated to oligonucleotides can be evaluated using the "G-less" transcript system of Sawadogo and Roeder [(1985) *Proc. Natl. Acad. Sci.* 82:4394-4398. While we do not wish to be limited to transcripts lacking guanosine residues, this assay allows screening of various moieties in a standardized manner. Those which are shown to be most active at preventing capping in this assay system are then incorporated into antisense oligonucleotides directed toward selected mRNA targets, such as ICAM-1 and CMV IE1/IE2. Briefly, this assay utilizes a HeLa cell extract for in vitro transcription of an exogenous DNA construct which encodes a G-less transcript. Such a transcript allows selective radiolabeling of capped mRNAs by the addition of [ $\alpha$ -P32]-GTP, a substrate for guanylyltransferase. Only capped mRNAs will be radiolabeled.

**Example 2**

**25 Inhibition of ICAM-1 mRNA capping**

The sequence of the ICAM-1 mRNA transcript is known up to and including the 5'-most nucleotide. Degitz, K., Lian-Jie, L., and Caughman, S.W. (1991) *J. Biol. Chem.* 266:14024-14030. The 5' end of the nascent mRNA target is shown in Table 1, with 5'-triphosphate included:

Table 1

ICAM-1 mRNA 5' end

5' pppGAGCUCCUCUGCUACUCAGA SEQ ID NO: 1

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An antisense oligonucleotide complementary to the 5'-most nucleotides of the ICAM-1 sequence in Table 1 is synthesized according to Example 1. This oligonucleotide (SEQ ID NO:2) is shown in Table 2. A series of oligonucleotides are synthesized having SEQ ID NO: 2 and comprising an interfering moiety. These compounds are also shown in Table 2. Both phosphodiester (P=O) and phosphorothioate (P=S) oligonucleotides are shown.

The first such modified compound is an oligonucleotide which comprises two extra nucleotides on its 3' end such that they overhang the 5' end of the nascent ICAM-1 mRNA target SEQ ID NO: 3. This compound is readily prepared using the methods of automated oligonucleotide synthesis described in Example 1. In the example shown, the overhanging nucleotides are AG, but other nucleotides can also be used.

The second compound comprises an oligonucleotide having SEQ ID NO: 2 having a triethylene tetramine (Tren) moiety conjugated to the 3' end. This compound is prepared as follows: The oligonucleotide having SEQ ID NO: 2 is synthesized as in Example 1. Oxidation of the 3' ribose residue of the oligonucleotide is achieved with 20 mM sodium periodate in 100 mM sodium acetate (pH 5.2) for 4 hours at 4°C. Oligonucleotide concentration is 0.4 mM. The reaction is quenched with glycerol. The product is precipitated from solution with ethanol and dried under vacuum. Conjugation of the triethylene tetramine to the oxidized 3' ribose residue is performed by incubating 0.4 mM oxidized oligonucleotide, 20 mM Tren, 200 mM sodium bicarbonate (pH 9.2) for three hours at room temperature. The enamine conjugation product is reduced by addition of greater than ten equivalents (relative to oligonucleotide) of solid sodium borohydride directly to the conjugation reaction, mixing and incubating for one hour at 4°C. The reaction solution is purified by standard ion exchange chromatography using a Pharmacia FPLC system.

The third compound comprises an oligonucleotide having SEQ ID NO: 2 having an MPG [N-(2-mercaptopropionyl)

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glycine:Cu(II)] coordination complex conjugated to the 3' end. This compound is prepared via postmodification procedures. The oligonucleotide is functionalized during automated DNA synthesis with a conjugation group, usually a primary amine, 5 sulfhydryl or ribose residue, at its 3' terminus. The MPG is derivatized with a complementary conjugation group, usually an aldehyde, carboxylic acid or primary amine. Sugiura, Y. and Hirayama, Y. (1976) *Inorg. Chem.* 15:679-682; Sugiura, Y., Hirayama, Y., Tanaka, H. and Ishizu, K., (1975) *J. Am. Chem.* 10 *Soc.* 97:5577-5581; Sugiura, Y., *Inorg. Chem.* 17:2176-2181. Conjugation is performed according to the postmodification procedures described by Uhlmann and Peyman [(1990) *Chem. Rev.* 90:543-584)].

The final compound shown comprises an oligonucleotide 15 having SEQ ID NO: 2 with a tetra peptide lys-his-gly-his conjugated to the 3' end. This compound is prepared as follows: The oligonucleotide is synthesized as in Example 1. Oxidation of the 3' ribose residue of the oligonucleotide is achieved with 20 mM sodium periodate in 100 mM sodium acetate (pH 5.2) 20 for 4 hours at 4°C. Oligonucleotide concentration is 0.4 mM. The reaction is quenched with glycerol. The product is precipitated from solution with ethanol and dried under vacuum. Conjugation of the peptide (N and C termini protected with Fmoc and an amide group, respectively) via the  $\epsilon$ -amino group of the 25 lysine residue to the aldehyde groups of the oxidized 3' ribose residue is performed by incubating 0.4 mM oxidized oligonucleotide, 20 mM peptide, 200 mM sodium bicarbonate (pH 9.2) for three hours at room temperature. The enamine conjugation product is reduced by addition of greater than ten 30 equivalents (relative to oligonucleotide) of solid sodium borohydride directly to the conjugation reaction, mixing and incubating for one hour at 4°C. The oligonucleotide-peptide conjugate is purified from the reducing reagent by size-exclusion chromatography (G10-120 Sephadex). The Fmoc amino 35 protecting group is removed in 1M triethylamine at room temperature in 30 minutes. The reaction solution is further purified by standard ion exchange chromatography using a

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Pharmacia FPLC system. Other peptides may generally be used as well.

**Table 2**  
**Oligonucleotides for interfering with**  
**capping of nascent ICAM-1 mRNA**

5	P=O	5' GTAGCAGAGGAGCTC	3'	SEQ ID NO: 2
	P=S	5' GTAGCAGAGGAGCTC	3'	SEQ ID NO: 2
	P=O (+ 2)	5' GTAGCAGAGGAGCTCAG	3'	SEQ ID NO: 3
	P=S (+ 2)	5' GTAGCAGAGGAGCTCAG	3'	SEQ ID NO: 3
10	P=O (+ Tren)	5' GTAGCAGAGGAGCTC-Tren	3'	SEQ ID NO: 2
	P=S (+ Tren)	5' GTAGCAGAGGAGCTC-Tren	3'	SEQ ID NO: 2
	P=O (+ MPG)	5' GTAGCAGAGGAGCTC-MPG:Cu(II)	3'	SEQ ID NO: 2
	P=S (+ MPG)	5' GTAGCAGAGGAGCTC-MPG:Cu(II)	3'	SEQ ID NO: 2
	P=O (+ peptide)	5' GTAGCAGAGGAGCTC-KHGH	3'	SEQ ID NO: 2
15	P=S (+ peptide)	5' GTAGCAGAGGAGCTC-KHGH	3'	SEQ ID NO: 2

### Example 3

**Ability of compounds to block capping of ICAM-1 mRNA determined by fingerprinting assay for presence of 5' cap**

A biochemical fingerprinting procedure for analysis of the 5' end of mRNA was originally used to define the chemical structure of the 5' cap [Cory, S. and Adams, J.M. (1975) *J. Mol. Biol.* 99:519-547]. This procedure is used here as an assay for the presence or absence of the methylated guanine cap structure. Briefly, the radiolabeled target mRNA is synthesized in vitro according to a generally used method such as that of Dignam et al. [(1983) *Nucl. Acids. Res.* 11:1475] in the presence of oligonucleotide compounds. The RNA is purified and digested with a mixture of pancreatic, T1 and T2 ribonucleases and fractionated by electrophoresis on cellulose acetate at pH 3.5, followed by electrophoresis on DEAE paper at pH 3.5. The

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presence of an electrophoretic spot corresponding to 7-methyl guanosine 5'-phosphate indicates the presence of a 5' cap.

#### Example 4

#### 5 Efficacy of compounds on blocking ICAM-1 expression in cell culture as determined by ELISA assay

The human lung carcinoma cell line A549 is obtained from the American Type Culture Collection (Bethesda MD). Cells are grown in Dulbecco's Modified Eagle's Medium (Irvine Scientific, Irvine CA) containing 1 gm glucose/liter and 10% fetal calf  
10 serum (Irvine Scientific). Human umbilical vein endothelial cells (HUVEC) (Clonetics, San Diego CA) are cultured in EGM-UV medium (Clonetics). HUVEC are used between the second and sixth passages. Human epidermal carcinoma A431 cells are obtained from the American Type Culture Collection and cultured  
15 in DMEM with 4.5 g/l glucose. Primary human keratinocytes are obtained from Clonetics and grown in KGM (Keratinocyte growth medium, Clonetics).

Cells grown in 96-well plates are washed three times with Opti-MEM (GIBCO, Grand Island, NY) prewarmed to 37°C. 100 µl  
20 of Opti-MEM containing either 10 µg/ml N-[1-(2,3-dioleyloxy)propyl]-N,N,N-trimethylammonium chloride (DOTMA, Bethesda Research Labs, Bethesda MD) in the case of HUVEC cells or 20 µg/ml DOTMA in the case of A549 cells is added to each well. Oligonucleotides are sterilized by centrifugation  
25 through 0.2 µm Centrex cellulose acetate filters (Schleicher and Schuell, Keene, NH). Oligonucleotides are added as 20x stock solution to the wells and incubated for 4 hours at 37°C. Medium is removed and replaced with 150 µl of the appropriate growth medium containing the indicated concentration of  
30 oligonucleotide. Cells are incubated for an additional 3 to 4 hours at 37°C then stimulated with the appropriate cytokine for 14 to 16 hours, as indicated. ICAM-1 expression is determined by ELISA assay: Cells are gently washed three times with a buffered isotonic solution containing calcium and magnesium  
35 such as Dulbecco's phosphate buffered saline (D-PBS). The cells are then directly fixed on the microtiter plate with 1 to

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2% paraformaldehyde diluted in D-PBS for 20 minutes at 25°C. The cells are washed again with D-PBS three times. Nonspecific binding sites on the microtiter plate are blocked with 2% bovine serum albumin in D-PBS for 1 hour at 37°C. Cells are  
5 incubated with ICAM-1 monoclonal antibody diluted in blocking solution for 1 hour at 37°C. Unbound antibody is removed by washing the cells three times with D-PBS. Antibody bound to the cells is detected by incubation with a 1:1000 dilution of biotinylated goat anti-mouse IgG (Bethesda Research  
10 Laboratories, Gaithersburg, MD) in blocking solution for 1 hour at 37°C. Cells are washed three times with D-PBS and then incubated with a 1:1000 dilution of streptavidin conjugated to  $\beta$ -galactosidase (Bethesda Research Laboratories) for 1 hour at 37°C. The cells are washed three times with D-PBS for 5  
15 minutes each. The amount of  $\beta$ -galactosidase bound to the ICAM-specific monoclonal antibody is determined by developing the plate in a solution of 3.3 mM chlorophenolred- $\beta$ -D-galactopyranoside, 50 mM sodium phosphate, 1.5 mM  $MgCl_2$ ; pH=7.2 for 2 to 15 minutes at 37°C. The concentration of the product  
20 is determined by measuring the absorbance at 575 nm in an ELISA microtiter plate reader.

#### Example 5

##### Inhibition of capping of cytomegalovirus IE1 and IE2 mRNA

The sequence of the CMV IE1/IE2 mRNA transcript is known.  
25 Stenberg, R.M., Witte, P.R. and Stinski, M.F. (1985) *J. Virol.* 56: 665-675; Stenberg, R.M., Thomsen, D.R., and Stinski, M.F. (1984) *J. Virol.* 49: 190-199. The 5' end of the nascent mRNA target shown in Table 3, with 5'-triphosphate included, is now known to terminate with a "G":

30

Table 3

CMV IE1/IE2 mRNA 5' end

5' pppGUCAGAUCCGCGGAGACGCC SEQ ID NO: 4

An antisense oligonucleotide complementary to the 5'-most nucleotides of the CMV IE1/IE2 sequence in Table 3 is

- 17 -

synthesized according to Example 1. This oligonucleotide (SEQ ID NO:4) is shown in Table 4. A series of oligonucleotides are synthesized having SEQ ID NO: 4 and comprising an interfering moiety. These compounds are also shown in Table 4. Both  
5 phosphodiester (P=O) and phosphorothioate (P=S) oligonucleotides are shown.

The first such compound is an oligonucleotide which comprises two extra nucleotides on its 3' end such that they overhang the 5' end of the nascent CMV IE1/IE2 mRNA target.  
10 This compound is readily prepared using the methods of automated oligonucleotide synthesis described in Example 1. In the example shown, the overhanging nucleotides are AG, but other nucleotides can also be used.

The second compound comprises an oligonucleotide having  
15 SEQ ID NO:4 having a triethylene tetramine (Tren) moiety conjugated to the 3' end. This compound is prepared as follows: oxidation of the 3' ribose residue of the oligonucleotide is achieved with 20 mM sodium periodate in 100 mM sodium acetate (pH 5.2) for 4 hours at 4°C. Oligonucleotide concentration is  
20 0.4 mM. The reaction is quenched with glycerol. The product is precipitated from solution with ethanol and dried under vacuum. Conjugation of the triethylene tetramine to the oxidized 3' ribose residue is performed by incubating 0.4 mM oxidized oligonucleotide, 20 mM Tren, 200 mM sodium bicarbonate (pH 9.2)  
25 for three hours at room temperature. The enamine conjugation product is reduced by addition of greater than ten equivalents (relative to oligonucleotide) of solid sodium borohydride directly to the conjugation reaction, mixing and incubating for one hour at 4°C. The reaction solution is purified by standard  
30 ion exchange chromatography using a Pharmacia FPLC system.

The third compound comprises an oligonucleotide having SEQ ID NO: 4 having an MPG [N-(2-mercaptopropionyl)glycine: Cu(II)] coordination complex conjugated to the 3' end. This compound is prepared via postmodification procedures. The  
35 oligonucleotide is functionalized during automated DNA synthesis with a conjugation group, usually a primary amine, sulfhydryl or ribose residue, at its 3' terminus. The MPG is

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derivatized with a complementary conjugation group, usually an aldehyde, carboxylic acid or primary amine. Sugiura, Y. and Hirayama, Y. (1976) *Inorg. Chem.* 15:679-682; Sugiura, Y., Hirayama, Y., Tanaka, H. and Ishizu, K., (1975) *J. Am. Chem. Soc.* 97:5577-5581; Sugiura, Y., *Inorg. Chem.* 17:2176-2181. Conjugation is performed according to the postmodification procedures described by Uhlmann and Peyman [(1990) *Chem. Rev.* 90:543-584)].

The final compound shown comprises an oligonucleotide having SEQ ID NO:4 having a tetra peptide lys-his-gly-his conjugated to the 3' end. This compound is prepared as follows: oxidation of the 3' ribose residue of the oligonucleotide is achieved with 20 mM sodium periodate in 100 mM sodium acetate (pH 5.2) for 4 hours at 4°C. Oligonucleotide concentration is 0.4 mM. The reaction is quenched with glycerol. The product is precipitated from solution with ethanol and dried under vacuum. Conjugation of the peptide (N and C termini protected with Fmoc and an amide group, respectively) via the  $\epsilon$ -amino group of the lysine residue to the aldehyde groups of the oxidized 3' ribose residue is performed by incubating 0.4 mM oxidized oligonucleotide, 20 mM peptide, 200 mM sodium bicarbonate (pH 9.2) for three hours at room temperature. The enamine conjugation product is reduced by addition of greater than ten equivalents (relative to oligonucleotide) of solid sodium borohydride directly to the conjugation reaction, mixing and incubating for one hour at 4°C. The oligonucleotide-peptide conjugate is purified from the reducing reagent by size-exclusion chromatography (G10-120 Sephadex). The Fmoc amino protecting group is removed in 1M triethylamine at room temperature in 30 minutes. The reaction solution is further purified by standard ion exchange chromatography using a Pharmacia FPLC system. Other peptides may also be used.

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Table 4

**Oligonucleotides for interfering with capping  
of nascent CMV IE1/IE2 mRNA**

	P=O	5' TCCAGGCGATCTGAC	3' SEQ ID NO: 5
5	P=S	5' TCCAGGCGATCTGAC	3' SEQ ID NO: 5
	P=O (+ 2)	5' TCCAGGCGATCTGAC-AG	3' SEQ ID NO: 6
	P=S (+ 2)	5' TCCAGGCGATCTGAC-AG	3' SEQ ID NO: 6
	P=O (+ Tren)	5' TCCAGGCGATCTGAC-Tren	3' SEQ ID NO: 5
	P=S (+ Tren)	5' TCCAGGCGATCTGAC-Tren	3' SEQ ID NO: 5
10	P=O (+ MPG)	5' TCCAGGCGATCTGAC-MPG:Cu(II)	3' SEQ ID NO: 5
	P=S (+ MPG)	5' TCCAGGCGATCTGAC-MPG:Cu(II)	3' SEQ ID NO: 5
	P=O (+ peptide)	5' TCCAGGCGATCTGAC-KHGH	3' SEQ ID NO: 5
	P=S (+ peptide)	5' TCCAGGCGATCTGAC-KHGH	3' SEQ ID NO: 5

**Example 6**

**15 Ability of compounds to block capping of ICAM-1 mRNA determined  
by fingerprinting assay for presence of 5' cap**

A biochemical fingerprinting procedure for analysis of the 5' end of mRNA was originally used to define the chemical structure of the 5' cap [Cory, S. and Adams, J.M. (1975) *J. Mol. Biol.* 99:519-547]. This procedure is used here as an assay for the presence or absence of the methylated guanine cap structure. Briefly, the radiolabeled target mRNA is synthesized in vitro according to a generally used method such as that of Dignam et al. [(1983) *Nucl. Acids. Res.* 11:1475]] in the presence of oligonucleotide compounds. The RNA is purified and digested with a mixture of pancreatic, T1 and T2 ribonucleases and fractionated by electrophoresis on cellulose acetate at pH 3.5, followed by electrophoresis on DEAE paper at pH 3.5. The presence of an electrophoretic spot corresponding to 7-methyl  
 25 guanosine 5'-phosphate indicates the presence of a 5' cap.  
 30

- 20 -

**Example 7**

**ELISA assay for inhibition of CMV replication by oligonucleotides which block capping of IE1/IE2 mRNA**

Oligonucleotides shown in Table 4 are tested for  
5 antiviral activity in an ELISA-based assay of CMV replication. Normal human dermal fibroblasts (Clonetics Corp., San Diego CA) are grown in serum-free medium (Clonetics) and used to seed 96-well plates. When cells are approximately 80% confluent, they are pretreated with compounds. Approximately 20 hours after  
10 pretreatment the medium (containing oligonucleotide) is carefully poured off and the cells washed twice with warmed fibroblast basal medium (FBM, Clonetics). Cells are then infected with 100  $\mu$ l/well of CMV stock diluted in FBM. The plates are incubated at 37°C for two hours. The medium  
15 (containing virus) is then carefully poured off and replaced with fresh, prewarmed FBM medium, 100 $\mu$ l per well. The plates are briefly incubated at 37°C and then 5  $\mu$ l of compound, diluted in FBM, is reintroduced into the medium in each well. Two days later, cells are post-treated again with  
20 oligonucleotides in the same way. On day six, the plates are prepared for ELISA.

In preparation for ELISA, the medium is carefully poured off the plates, and cells are fixed in 200  $\mu$ l of absolute ethanol per well. Cells are fixed for 30 minutes at room temperature,  
25 then ethanol is removed and plates are air-dried. Plates are blocked for one hour prior to ELISA with PBS containing 2% BSA. Blocking solution is removed and 100  $\mu$ l of an anti-CMV antibody, diluted 1:2000 in PBS with 1% BSA, is added. Cells are incubated in antibody for one hour at 37°C and washed three  
30 times in PBS. The secondary antibody, biotinylated goat anti-mouse IgG (Bethesda Research Labs, MD), is diluted 1:1000 in PBS with 1% BSA, and incubated with cells for one hour at 37°C. Cells are then washed and incubated for one hour at 37°C in streptavidin-B-D-galactosidase. Color is developed with  
35 chlorophenol red-B-D-galactopyranoside, 20 mg dissolved in 10 ml of 50 mM Na Phosphate, 1.5 mM MgCl<sub>2</sub>; plates are shaken for 10 minutes and the absorbance is read at 575 nm.

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## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

(i) APPLICANT: ISIS Pharmaceuticals and  
Brenda Baker

5 (ii) TITLE OF INVENTION: Compositions and Methods for  
Modulating

RNA Activity Through Interference with Capping of  
Nascent RNA

(iii) NUMBER OF SEQUENCES: 6

10 (iv) CORRESPONDENCE ADDRESS:

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15 (D) STATE: PA

(E) COUNTRY: USA

(F) ZIP: 19103

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: DISKETTE, 3.5 INCH, 1.44 Mb STORAGE

20 (B) COMPUTER: IBM PS/2

(C) OPERATING SYSTEM: PC-DOS

(D) SOFTWARE: WORDPERFECT 5.0

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: n/a

25 (B) FILING DATE: herewith

(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 847,054

(B) FILING DATE: 03-APR-1992

30 (vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: PCT/US91/03636

(B) FILING DATE: 22-MAY-1991

(vii) PRIOR APPLICATION DATA:

- 22 -

(A) APPLICATION NUMBER: 527,559

(B) FILING DATE: 23-MAY-1990

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5 (B) REGISTRATION NUMBER: 28,937

(C) REFERENCE/DOCKET NUMBER: ISIS-1120

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10 (2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

15 (D) TOPOLOGY: unknown

(iv) ANTISENSE: no

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

GAGCUCCUCU GCUACUCAGA (20)

(2) INFORMATION FOR SEQ ID NO: 2:

20 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: unknown

25 (iv) ANTISENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GTAGCAGAGG AGCTC (15)

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 17

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: unknown

(iv) ANTISENSE: yes

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GTAGCAGAGG AGCTCAG (17)

(2) INFORMATION FOR SEQ ID NO: 4:

- 23 -

- (i) SEQUENCE CHARACTERISTICS:  
    (A) LENGTH: 21  
    (B) TYPE: nucleic acid  
    (C) STRANDEDNESS: single  
5     (D) TOPOLOGY: unknown  
    (iv) ANTISENSE: no  
    (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:  
        GUCAGAUCCG CUGGAGACGC C (21)
- (2) INFORMATION FOR SEQ ID NO: 5:  
10     (i) SEQUENCE CHARACTERISTICS:  
        (A) LENGTH: 15  
        (B) TYPE: nucleic acid  
        (C) STRANDEDNESS: single  
        (D) TOPOLOGY: unknown  
15     (iv) ANTISENSE: yes  
        (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:  
            TCCAGGCGAT CTGAC (15)
- (2) INFORMATION FOR SEQ ID NO: 6:  
    (i) SEQUENCE CHARACTERISTICS:  
20     (A) LENGTH: 17  
        (B) TYPE: nucleic acid  
        (C) STRANDEDNESS: single  
        (D) TOPOLOGY: unknown  
    (iv) ANTISENSE: yes  
25     (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:  
        TCCAGGCGAT CTGACAG (17)

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## CLAIMS

What is claimed is:

1. An oligonucleotide for inhibiting the capping of a preselected nascent mRNA which is specifically hybridizable  
5 with said nascent mRNA.
2. The oligonucleotide of claim 1 which is specifically hybridizable with a 5' region of said nascent mRNA.
3. The oligonucleotide of claim 1 which is 10 to 25 nucleotides in length.
- 10 4. The oligonucleotide of claim 1 wherein the nascent mRNA is from about 20 to about 80 nucleotides in length.
5. The oligonucleotide of claim 1 which is capable of sterically interfering with capping of the nascent mRNA.
6. The oligonucleotide of claim 1 which comprises at least  
15 one amino acid, polyamine, coordination complex of a metal, metal ion or macrocycle at the 3' end of the oligonucleotide.
7. The oligonucleotide of claim 6 which comprises at least one nucleotide at the 3' end which extends beyond the nucleotide units of the 5' portion of the nascent mRNA.
- 20 8. The oligonucleotide of claim 1 which is capable of chemically modifying the nascent mRNA.
9. The oligonucleotide of claim 8 wherein the chemical modification is dephosphorylation of the 5'-triphosphate or 5'-diphosphate of the nascent mRNA.
- 25 10. The oligonucleotide of claim 9 wherein the dephosphorylation yields a 5'-monophosphate or a hydroxyl group.

- 25 -

11. The oligonucleotide of claim 1 wherein the capping enzyme is a guanylyltransferase.
12. A method of inhibiting capping of a preselected nascent mRNA comprising contacting said nascent mRNA or cells, tissue  
5 or a bodily fluid containing said nascent mRNA with an oligonucleotide which is specifically hybridizable with said nascent mRNA.
13. The method of claim 12 wherein said specific hybridization of the oligonucleotide is with a 5' region of  
10 said nascent mRNA.
14. The method of claim 12 wherein the oligonucleotide is 12 to 25 nucleotides in length.
15. The method of claim 12 wherein the nascent mRNA is from about 20 to about 80 nucleotides in length.
16. The method of claim 12 wherein the nascent mRNA is rendered inaccessible to the capping enzyme by steric interference.
17. The method of claim 12 wherein the oligonucleotide  
20 comprises at least one amino acid, polyamine, coordination complex of a metal, metal ion or macrocycle at the 3' end of the oligonucleotide.
18. The method of claim 17 wherein the oligonucleotide comprises at least one nucleotide at the 3' end which extends  
25 beyond the nucleotide units of the 5' portion of the nascent mRNA.
19. The method of claim 12 wherein the inhibition comprises chemically modifying the nascent mRNA.
20. The method of claim 19 wherein the chemical modification

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is dephosphorylation of the 5'-triphosphate or 5'-diphosphate of the nascent mRNA.

21. The method of claim 20 wherein the dephosphorylation yields a 5'-monophosphate or a hydroxyl group.

5 22. The method of claim 12 wherein said capping enzyme is a guanylyltransferase.

23. A method of interfering with the function of an mRNA by inhibiting the capping of said mRNA in its nascent form according to the method of claim 12.

10 24. The method of claim 23 wherein the function of the mRNA to be interfered with is export of the mRNA from the nucleus.

25. The method of claim 23 wherein the function of the specific mRNA to be interfered with is translation of the mRNA.

15 26. The method of claim 23 wherein the function of the specific mRNA to be interfered with is stability of the mRNA.

27. The method of claim 23 wherein the mRNA to be interfered with is expressed in a cell-, tissue- or disease-specific manner.

## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US94/03401

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : A61K 48/00, 31/70, 31/74, 31/795; C07H 21/04

US CL : 514/44; 536/24.5

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/44; 536/24.5

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS

search terms: antisense, oligo?, cap

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X --- Y	Biomedica et Biochimica Acta, Volume 48, Number 1, issued 1989, Westermann et al., "Inhibition of Expression of SV40 Virus Large T-Antigen by Antisense Oligodeoxyribonucleotides", pages 85-93, especially pages 90-91.	1-6, 11-17, 22, 23, 25, 27 ----- 7-10, 18-21, 24, 26
Y	Cell, Volume 63, issued 05 October 1990, J. Hamm et al., "Monomethylated Cap Structures Facilitate RNA Export From The Nucleus", pages 109-118, especially page 112.	24, 26
Y	Molecular Biology of Medicine, Volume 5, issued 1988, J. Ross, "Messenger RNA Turnover In Eukaryotic Cells", pages 1-14, especially page 7.	24, 26



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	
*A* document defining the general state of the art which is not considered to be of particular relevance	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
*E* earlier document published on or after the international filing date	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
*L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
*O* document referring to an oral disclosure, use, exhibition or other means	*Z* document member of the same patent family
*P* document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

16 JUNE 1994

Date of mailing of the international search report

05 JUL 1994

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## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US94/03401

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Annual Review of Biochemistry, Volume 54, issued 1985, K. Moldave, "Eukaryotic Protein Synthesis", pages 1109-1149, especially pages 1121-1125.	7-10, 18-21
Y	Chemical Reviews, Volume 90, No. 4, issued June 1990, E. Uhlmann et al., "Antisense Oligonucleotides: A New Therapeutic Principle", pages 543-584, especially pages 546-558.	7-10, 18-21
Y	Journal of the American Chemical Society, Volume 113, issued 1991, A. Modak et al., "Toward Chemical Ribonucleases. 2. Synthesis and Characterization of Nucleoside-Bipyridine Conjugates. Hydrolytic Cleavage of RNA By Their Copper(II) Complexes", pages 283-291, especially pages 287-288.	8, 19
Y	Gene, Volume 61, issued 1987, P. Verspieren et al., "An Acridine-Linked Oligodeoxynucleotide Targeted To The Common 5' End Of Trypanosome mRNAs Kills Cultured Parasites", pages 307-315, especially pages 309-311.	8, 19
Y	Journal of the American Chemical Society, Volume 111, J. Chin et al., "Co(III) Complex Promoted Hydrolysis of Phosphate Diesters: Comparison in Reactivity of Rigid cis-Diaquotetraazacobalt(III) Complexes", pages 186-190, especially page 188.	9, 10, 20, 21
Y	D. Apirion, "PROCESSING OF RNA", published 1984 by CRC Press, Inc. (Boca Raton, FL), pages 119-132, see pages 121-124.	9, 10, 20, 21